FULL PAPER

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Mononuclear Co(III), Ni(II) and Cu(II) complexes of 2-(2,4dichlorobenzamido)‐N'‐(3,5‐di‐tert‐butyl‐2‐ hydroxybenzylidene)benzohydrazide: Structural insight and biological assay

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A series of mononuclear metal complexes of Co(III), Ni(II) and Cu(II) with 2‐ (2,4‐dichlorobenzamido)‐N′‐(3,5‐di‐t*ert‐*butyl‐2‐hydroxybenzylidene)benzohydrazide A series of mononuclear metal complexes of Co(III), Ni(II) and Cu(II) with 2-
(2,4-dichlorobenzamido)-*N'*-(3,5-di-*tert*-butyl-2-hydroxybenzylidene)benzohydrazide
(LH₃) have been synthesized and characterized using vari (2,4-dichlorobenzamido)- N' -(3,5-di-tert-butyl-2-hydroxybenzylidene)benzohydrazide
(LH_3) have been synthesized and characterized using various physico-chemical,
spectroscopic and single crystal X-ray diffraction techn ies of $[Co(LH)(LH₂)]^T$ ₁^O (4) revealed the presence of both amido and imidol tautomeric forms of LH_3 , resulting in a distorted octahedral geometry around the Co(III) ion. $[Ni(\mathbf{L}\mathbf{H})(H_2O)]\cdot H_2O$ (5) and $[Cu(\mathbf{L}\mathbf{H})(H_2O)]\cdot H_2O$ (6) are isomorphous structures and crystallize in the monoclinic $P2_1/c$ space
group. The crystal structures of **4, 5** and **6** are stabilized by hydrogen bonds
formed by the enclathrated water molecules, C-H $\cdots \pi$ and $\pi \cdots$ group. The crystal structures of 4, 5 and 6 are stabilized by hydrogen bonds Complexes along with the ligand (LH_3) were screened for their in vivo formed by the enclathrated water molecules, C-H \cdots π and $\pi \cdots \pi$ interactions. in vitro antioxidant activity (DPPH free radical scavenging assay). Metal anti-inflammatory activity (carrageenan-induced rat paw edema method) and *in vitro* antioxidant activity (DPPH free radical scavenging assay). Metal complexes have shown significant anti-inflammatory and antioxidant poten

KEYWORDS

anti-inflammatory activity, antioxidant activity, crystal structure, di-tert-butyl phenylhydrazone, isomorphous metal complexes

1 | INTRODUCTION

Oxidation processes are requisite for life; however, they can have toxic potential effects as they generate excess free radicals that can cause oxidative damage to proteins, membranes and genes. $[1-3]$ These reactive free radicals have been associated as mediators in an assortment of many diseases, such as arthritis, inflammation, cancer, neurological and heart diseases.[1,4–6] The reactive oxygen species generated by infiltrated neutrophils act as venomspecies generated by infiltrated neutrophils act as venom-
ous agents that are involved in the cyclooxygenase-2 species generated by infiltrated neutrophils act as venom-
ous agents that are involved in the cyclooxygenase-2
(COX-2) and 5-lipoxygenase (5-LOX) mediated conversion of arachidonic acid into proinflammatory intermedi-(COX-2) and 5-lipoxygenase (5-LOX) mediated conversion of arachidonic acid into proinflammatory intermediates, hence, enhancing the inflammatory process.^[7–9] Therefore, the compounds with antioxidant/radical scavenging properties could be anticipated to offer protection in arthritis and inflammation. Many studies on commerreflection, the compounds with antioxidant/radical scavenging properties could be anticipated to offer protection
in arthritis and inflammation. Many studies on commer-
cially available nonsteroidal anti-inflammatory drugs

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and their metal complexes have proven their simultaneous free radical scavenging properties.^[10,11]

Aroylhydrazones (AHs) are an elite class of compounds that have drawn much attention as versatile ligands with a variety of coordination modes with transition metals.^[12] AHs exhibit amido-imidol tautomerism in solution and coordinate to metal ions through its amido/imidol form. In exceptional cases, both forms simultaneously coordinate to the metal ion. The possibility of tautomerism in this class of compounds has proved attractive in the field of pharmacology. Most of the first row transition metals either alone or in their complex form are biologically important with a number of bioactivities. Transition metal complexes of AHs present a diverse range of biological applications, such as anti-inflammatory, antioxidant, antiproliferative and antitumor activities.^[13-15] These transition metal complexes have shown greater biological potential than the proligands. In addition, transition metal complexes of AHs are known to act as efficient and selective catalysts towards various chemical reactions.^[16,17] Hydrazones complexes of Aris are known to act
efficient and selective catalysts towards various
mical reactions.^[16,17]
Hydrazones containing di-*tert*-butyl phenol moiety as

a key structural feature are of conspicuous interest, due to their diverse antioxidant, anticancer, antifungal, antia key structural feature are of conspicuous interest, due
to their diverse antioxidant, anticancer, antifungal, anti-
bacterial and dual COX/LOX inhibitory activities.^[18–20]
An array of metal complexes of di-*tert*-buty phenylhydrazones has been described in the litera-An array of metal complexes of di-*tert*-butyl phenylhydrazones has been described in the literature.^[16,17,21–24] Many of them did not divulge a biological relevance. Based on the above facts, our present endeavor emphasizes the synthesis, structural characterization, Fix any of them and not divulge a biological
relevance. Based on the above facts, our present endeavor
emphasizes the synthesis, structural characterization,
antioxidant and anti-inflammatory activities of transition relevance. Based on the above facts, our present endeavor
emphasizes the synthesis, structural characterization,
antioxidant and anti-inflammatory activities of transition
metal complexes of newly synthesized tridentate di butyl phenylhydrazone (LH_3) derived from the condensaantioxidant and anti-innaminatory activities of transition
metal complexes of newly synthesized tridentate di-*tert*-
butyl phenylhydrazone (LH_3) derived from the condensa-
tion of 2-(2,4-dichlorobenzamido)benzohydrazi metal complexes of hewly synthesized
butyl phenylhydrazone (**LH**₃) derived fro
tion of 2-(2,4-dichlorobenzamido)benz
3,5-di-*tert*-butyl-2-hydroxybenzaldehyde.

2 | EXPERIMENTAL

2.1 | Materials and physical measurements

All the reagents and solvents were purchased commercially and used without further purification. Elemental analysis (C, H, N) was performed on Thermoquest CHN analyzer. The metal content of the complexes was determined according to the literature procedure.^[25] ¹H and analysis (C, H, N) was performed on Thermoquest CHN
analyzer. The metal content of the complexes was deter-
mined according to the literature procedure.^[25] ¹H and
¹³C NMR spectra were measured on Bruker AV-400 and analyzer. The metal content of the complexes was deter-
mined according to the literature procedure.^[25] ¹H and
¹³C NMR spectra were measured on Bruker AV-400 and
AGILENT VNMRS-400 spectrometer, respectively, in ¹³C NMR spectra were measured on Bruker AV-400 and AGILENT VNMRS-400 spectrometer, respectively, in an internal standard. Infrared (IR) spectra were recorded dimethylsulfoxide (DMSO)- d_6 with tetramethylsilane as
an internal standard. Infrared (IR) spectra were recorded
on a Nicolet-6700 FT-IR spectrometer using KBr discs in an internal standard. Infrared (IR) spectra were recorded
on a Nicolet-6700 FT-IR spectrometer using KBr discs in
the 4000–400 cm⁻¹ region. Absorbance spectra were

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recorded on a JASCO V-670 UV–Vis spectrophotometer. CHIMMALAGI ET AL.

Fecorded on a JASCO V-670 UV–Vis spectrophotometer.

ESI-MS were recorded on a Waters XEVO TQS micromass spectrometer. EPR spectra were recorded at both recorded on a JASCO v-670 Ov-vis spectrophotometer.
ESI-MS were recorded on a Waters XEVO TQS micro-
mass spectrometer. EPR spectra were recorded at both
room temperature and 77 K on a Varian E-4 X-band spectrometer. Conductance measurements of complexes mass spectrometer. EFR spectra were recorded at both
room temperature and 77 K on a Varian E-4 X-band spec-
trometer. Conductance measurements of complexes
(50 μ M) were recorded in DMF using an ELICO-CM-82 conductivity bridge. Thermogravimetric analyses (TGA) (50 μ M) were recorded in DMF using an ELICO-CM-82 conductivity bridge. Thermogravimetric analyses (TGA) were carried out over the temperature range of 25– 1000°C using Universal V4.5A TA instrument.

2.2 [|] Synthesis of 2‐(2,4‐ dichlorobenzamido)‐N′‐(3,5‐di‐tert‐butyl‐2‐ hydroxybenzylidene)benzohydrazide (LH3)

LH₃ was prepared in three steps as reported earlier in a **EH₃** was prepared in times steps as reported earlier in a similar case.^[26] In the first step, methyl anthranilate (1; 3.02 g, 20 mmol) was suspended in benzene (300 ml), to which 2,4-dichlorobenzoyl chloride (4.16 g 3.02 g, 20 mmol) was suspended in benzene (300 ml), to was added and stirred for 3 h at room temperature to generate methyl $2(2,4-{\text{dichlorobenzoyl}}{\text{chlorobenzoyl}})$ chloride (4.16 g, 20 mmol)
was added and stirred for 3 h at room temperature to generate methyl $2-(2,4-{\text{dichlorobenzamido)}}$ benzoate (2; yield: 91%), which was then hydrazinolyzed at reflux temwas added and stirted for 5 if at foom temperature to generate
erate methyl 2-(2,4-dichlorobenzamido)benzoate (2;
yield: 91%), which was then hydrazinolyzed at reflux tem-
perature to produce 2-(2,4-dichlorobenzamido) benzohydrazide (3; yield: 76%). Finally, the methanolic solution of 3,5–di-*tert*-butyl-2-hydroxybenzamido)
solution of 3,5–di-*tert*-butyl-2-hydroxybenzaldehyde (2.34 g, 10 mmol) was added to a methanolic suspension of 3 (3.24 g, 10 mmol). The reaction mixture along with a catalytic amount of glacial acetic acid was refluxed for 3 h. Progress of the reaction was monitored by TLC. The white precipitate formed was filtered off, washed with hot methanol and dried in air. Schematic representation for the synthesis of $LH₃$ is shown in Scheme 1.

LH₃: color: colorless. Isolated yield: 84%. m.p. 250-252°C. Anal. calcd for $C_{29}H_{31}Cl_2N_3O_3$: C, 64.44; H, 5.78; N, 7.77%. Found: C, 64.70; H, 5.53; N, 7.68%. IR (ν, KBr, **CH₃:** Color: Coloriess. Isolated yield: 84%. In.p. 250–252°C. Anal. calcd for C₂₉H₃₁Cl₂N₃O₃: C, 64.44; H, 5.78; N, 7.77%. Found: C, 64.70; H, 5.53; N, 7.68%. IR (v, KBr, cm⁻¹): 3444, 3326, 3232, 1680, 1660, 252 C. Anal. calcu for C₂₉H₃₁C₁₂N₃O₃. C, 64.44, H, 5.78,
N, 7.77%. Found: C, 64.70; H, 5.53; N, 7.68%. IR (v, KBr,
cm⁻¹): 3444, 3326, 3232, 1680, 1660, 1607. ¹H-NMR
(400 MHz, DMSO-d₆, δ, ppm): 1.28 (9H, s, (9H, s, *tert*–Bu), 7.20 (1H, d, C23H, J = 2 Hz), 7.31 (1H, d, C23H, J (400 MHz, DMSO-d₆, δ , ppm): 1.28 (9H, s, tert-Bu), 1.39 (9H, s, tert-Bu), 7.20 (1H, d, C23H, J = 2 Hz), 7.31 (1H, d, C29H, J = 2 Hz), 7.34 (1H, t, C11H, J = 8 Hz), 7.65– 7.58 (2H, m, C4H and C10H), 7.70 (1H, d, C5H, $J = 8$) Hz), 7.76 (1H, d, C2H, $J = 1.6$ Hz), 7.81 (1H, d, C9H, J $= 7.6$ Hz), 8.25 (1H, d, C12H, J $= 8$ Hz), 8.52 (1H, s, C15H), 11.13 (1H, s, O3H), 12.16 (1H, s, N2H), 12.33 (1H, 3, 1.76 (1H, 3, C2H, 3 = 1.6 Hz), 7.81 (1H, 3, C9H, 3

(1H, 8, 225 (1H, 4, C12H, 3 = 8 Hz), 8.52 (1H, s, C15H), 11.13 (1H, s, O3H), 12.16 (1H, s, N2H), 12.33

(1H, s, N1H). ¹³C-NMR (100 MHz, DMSO-d₆, δ , ppm): $= 7.6$ Hz), 8.25 (1H, d, C12H, $J = 8$ Hz), 8.52 (1H, s, C15H), 11.13 (1H, s, O3H), 12.16 (1H, s, N2H), 12.33
(1H, s, N1H). ¹³C-NMR (100 MHz, DMSO-d₆, δ , ppm):
29.77 ((CH₃)₃), 31.74 ((CH₃)₃), 34.34 (C-(CH₃ (C_{15H)}, T_{1.13} (T_H, s, O_{5H)}, 12.16 (TH, s, N_{2H)}, 12.33
(1H, s, N1H). ¹³C-NMR (100 MHz, DMSO-d₆, δ , ppm):
29.77 ((CH₃)₃), 31.74 ((CH₃)₃), 34.34 (C-(CH₃)₃), 35.09
(C-(CH₃)₃), 117.35 (C16), 12 128.23 (C4), 130.10 (C5), 130.84 (C2), 131.73 (C6), 29.77 ((CH₃)₃), 31.74 ((CH₃)₃), 34.34 (C-(CH₃)₃), 35.09
(C-(CH₃)₃), 117.35 (C16), 124.66 (C29), 126.28 (C23),
128.23 (C4), 130.10 (C5), 130.84 (C2), 131.73 (C6),
136.20, 122.73, 132.79, 126.28, 129.15, 123. matic), 135.24 (C1), 135.92 (C18), 138.12 (C3), 140.99 (C24), 152.37 (C15), 155.18 (C17), 164.19 (C14), 164.32 136.20, 122.73, 132.79, 126.28, 129.15, 123.16 (C8-13, aromatic), 135.24 (C1), 135.92 (C18), 138.12 (C3), 140.99 (C24), 152.37 (C15), 155.18 (C17), 164.19 (C14), 164.32 (C7). ESI-MS (m/z): 540 $\left[\mathbf{L}\mathbf{H}_3 + \mathbf{H}\right]^+$. (ε/dm3 mol−¹ cm−¹): 266 (38 846), 299 nm (55 756).

2.3 | Syntheses of metal complexes

2.3.1 | $[Co(LH)(LH₂)]·H₂O(4)$

A methanolic solution of $Co(CH_3COO)_{2} \cdot 4H_{2}O$ (0.062 g, 0.25 mmol) was added dropwise to a methanolic suspension of LH_3 (0.270 g, 0.5 mmol). The reaction mixture was then refluxed on a water bath for 4 h (Scheme 2a). The resultant solution was left to evaporate until reddish brown crystals of [Cu(LH)(H₂O)]H₂O suitable for X-ray diffraction studies were separated.

Color: reddish brown. Isolated yield: 76%. Anal. calcd for $C_{58}H_{61}Cl_4CoN_6O_7$: C, 60.32; H, 5.32; N, 7.28; Co, 5.10%. Found: C, 59.98; H, 5.14; N, 7.37; Co, 5.02%. IR Colof: Feddish brown: Isolated yield: 76%. Anal. card
for C₅₈H₆₁Cl₄CoN₆O₇: C, 60.32; H, 5.32; N, 7.28; Co,
5.10%. Found: C, 59.98; H, 5.14; N, 7.37; Co, 5.02%. IR
(ν, KBr, cm⁻¹): 3423, 3347, 1681, 1612, 1585, MS (M) $C_{58}H_{61}Cl_4CON_6O_7$. C, 60.52, H, 5.52, N, 7.28, Co, 5.02%. IR
5.10%. Found: C, 59.98; H, 5.14; N, 7.37; Co, 5.02%. IR
(v, KBr, cm⁻¹): 3423, 3347, 1681, 1612, 1585, 1320. ESI-
MS (m/z): 1135 [Co(LH)(LH₂) + H nm (ε/dm³ mol⁻¹ cm⁻¹): 268 (79 700), 322 (30 915), 442 nm (21 348). Molar conductance $(\Omega^{-1} \text{ cm}^2 \text{ mol}^{-1})$: 2.52.

2.3.2 | $[Ni(LH)(H₂O)] \cdot H₂O(5)$

A mixture of LH_3 (0.270 g, 0.5 mmol), sodium acetate $(0.082 \text{ g}, 2.0 \text{ mmol})$ and NiCl₂·6H₂O $(0.119 \text{ g}, 0.5 \text{ mmol})$ in methanol was refluxed on a water bath for 4 h (Scheme 2b), and the resultant solution was left to evaporate until reddish brown crystals of [Ni(LH) $(H₂O)|·H₂O$ suitable for X-ray diffraction studies were separated.

Color: reddish brown. Isolated yield: 69%. Anal. calcd for $C_{29}H_{33}Cl_2N_3NiO_5$: C, 55.01; H, 5.25; N, 6.64; Ni, 9.27%. Found: C, 55.32; H, 5.17; Ni, 9.16; N, 6.52%. IR (ν, KBr, cm−¹): 3625, 3559, 3431, 1679, 1613, 1594, 1324. ¹H-NMR (400 MHz, CDCl₃, δ , ppm): 0.90 (9H, s, tert-01 C₂₉H₃₃C₁₂N₃N₁O₅: C, 55.01, H, 5.25, N, 6.64, N1,
0.27%. Found: C, 55.32; H, 5.17; Ni, 9.16; N, 6.52%. IR
ν, KBr, cm⁻¹): 3625, 3559, 3431, 1679, 1613, 1594, 1324.
H-NMR (400 MHz, CDCl₃, δ, ppm): 0.90 (9H 9.27%. Found: C, 55.52, H, 5.17, NI, 9.16, N, 6.52%. IK
(v, KBr, cm⁻¹): 3625, 3559, 3431, 1679, 1613, 1594, 1324.
¹H-NMR (400 MHz, CDCl₃, δ , ppm): 0.90 (9H, s, tert-
Bu), 1.21 (9H, s, tert-Bu), 6.73–7.04 (4H, m, Bu), 1.21 (9H, s, tert-Bu), 6.73-7.04 (4H, m, C23H, C29H, C10H and C11H), 7.27-7.47 (3H, m, C2H, C4H and C5H,) 8.19 (2H, d, C12H and C9H, $J = 7.2$ Hz), 8.69 (1H, s, C15H), 9.67 (1H, s, N1H). 13C‐NMR (100 MHz, CDCl₃, δ , ppm): 29.80 ((CH₃)₃), 30.79 ((CH₃)₃), 33.60

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(C-(CH₃)₃), 35.25 (C-(CH₃)₃), 117.14 (C16), 121.23 (C29), 126.33 (C23), 127.84 (C4), 129.53 (C5), 130.79 (C2 and Chemistry

(C-(CH₃)₃), 35.25 (C-(CH₃)₃), 117.14 (C16), 121.23 (C29),

126.33 (C23), 127.84 (C4), 129.53 (C5), 130.79 (C2 and

C6), 138.69, 120.13, 131.62, 124.12, 128.90, 136.53 (C8– 13, aromatic), 132.25 (C1), 133.61 (C18), 137.75 (C3), 139.82 (C24), 153.12 (C17), 157.83 (C15), 162.06 (C7), 172.69, 120.13, 131.62, 124.12, 126.90, 136.53 (C₆–13, aromatic), 132.25 (C1), 133.61 (C18), 137.75 (C3), 139.82 (C24), 153.12 (C17), 157.83 (C15), 162.06 (C7), 172.69 (C14). ESI-MS (m/z): 613 [Ni(LH)(NH₃) + H]⁺. 172.69 (C14). ESI-MS (m/z): 613 [Ni(**LH**)(NH₃) + H]⁺.
UV–Vis: $\lambda_{\text{max}}/\text{nm}$ (ε/dm^3 mol⁻¹ cm⁻¹): 267 (67 759), 320
(34 476), 428 nm (36 290). Molar conductance UV–Vis: $\lambda_{\text{max}}/\text{nm}$ (ε/dm^3 mol⁻¹ cm⁻¹): 267 (67 759), 320 (34 476), 428 nm (36 290). Molar conductance $(\Omega^{-1} \text{ cm}^2 \text{ mol}^{-1})$: 4.91.

SCHEME 2 (a) Synthetic route for the preparation of [Co(LH) $(LH₂)$]·H₂O (4). (b) Synthetic route for the preparation of [Ni(LH) $(H_2O)|H_2O(5)$ and $[Cu(LH)(H_2O)]H_2O(6)$

Reaction conditions: i) Benzene, R.T, 3 h. ii) N₂H₄.H₂O, MeOH, Reflux, 4 h. iii) 3,5-di-tert-butyl-2hydroxybenzaldehye, MeOH, AcOH, Reflux, 3 h.

SCHEME 1 Synthetic route for the preparation of LH₃

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2.3.3 | $\text{[Cu(LH)(H₂O)]·H₂O (6)}$

A methanolic solution of $Cu(CH_3COO)_{2}·H_2O$ (0.10 g, 0.5 mmol) was added dropwise to a methanolic suspension of LH_3 (0.270 g, 0.5 mmol). The reaction mixture was then refluxed on a water bath for 4 h (Scheme 2b). Single crysthe tals of [Cu(LH)(H₂O)]H₂O suitable for X-ray diffraction
tals of [Cu(LH)(H₂O)]H₂O suitable for X-ray diffraction studies were obtained on slow evaporation of the resultant solution at ambient temperature.

Color: green. Isolated yield: 74%. Anal. calcd for $C_{29}H_{33}Cl_2CuN_3O_5$: C, 54.59; H, 5.21; Cu, 9.96; N, 6.59%. Found: C, 55.01; H, 5.23; Cu, 9.73; N, 6.45. IR (ν, KBr, cm⁻¹): 3644, 3553, 3442, 1677, 1611, 1589, 1318. ESI-MS 0101: green. Isolated yield. 74%. Aflal. calcd 101
33Cl₂CuN₃O₅: C, 54.59; H, 5.21; Cu, 9.96; N, 6.59%.
d: C, 55.01; H, 5.23; Cu, 9.73; N, 6.45. IR (v, KBr,
): 3644, 3553, 3442, 1677, 1611, 1589, 1318. ESI-MS (m/z): 618 $\text{[Cu(LH)(NH_3) + H]}^+$. UV–Vis: $\lambda_{\text{max}}/ \text{nm}$ $(\varepsilon/\text{dm}^3 \text{ mol}^{-1} \text{ cm}^{-1})$: 267 (67 493), 323 (40 145), 416 (37 988), 666 nm (612). Molar conductance (Ω^{-1} cm² mol⁻¹): 4.79.

2.4 [|] Single crystal X‐ray diffraction studies

Single crystal data were collected at 100 K on a Rigaku **studies**
Single crystal data were collected at 100 K on a Rigaku
SuperNova, Dualflex, AtlasS2 diffractometer using Cu-Kα radiation ($\lambda = 1.54184$ Å). CrysAlis Pro software was used for data collection, absorption correction and data reduction.^[27] The structures were solved by a direct Example 1.34184 A). CrysAlls Fro soliware was
used for data collection, absorption correction and data
reduction.^[27] The structures were solved by a direct
method with SHELXD-2014/6 and refined using dised for data conection, absorption correction and data
reduction.^[27] The structures were solved by a direct
method with SHELXD-2014/6 and refined using
SHELXL-2014/6 program package.^[28] The structure of **4** method with SHELXD-2014/6 and refined using SHELXL-2014/6 program package.^[28] The structure of **4** was pseudo-centrosymmetric and could be solved in the space group Pbca; however, refinement in this setting has failed. As a consequence of the psuedosymmetry, was pseudo-centrosymmetric and could be solved in the
space group *Pbca*; however, refinement in this setting
has failed. As a consequence of the psuedosymmetry,
the overall structure was refined as a two-component inversion twin with a final Flack parameter of 0.273(4). Hydrogen atom positions were calculated geometrically and refined using the riding model. Mercury CSD 2.0 pro $gram^{[29]}$ was used for molecular graphics. The structural data have been deposited with the Cambridge Crystallographic Data Centre (CCDC) with reference numbers 1822746, 1822747 and 1822748 for 4, 5 and 6, respectively.

2.5 | DPPH radical scavenging activity

The antioxidant activity of the titled compounds was measured on the basis of the free radical scavenging The antioxidant activity of the titled compounds was
measured on the basis of the free radical scavenging
activity by the 2,2-diphenyl-1-picrylhydrazyl (DPPH) method.^[30] The stock solution of DPPH was prepared by dissolving 3.9432 mg DPPH in 100 ml of methanol (0.1 mM) and stored at 4°C until use; 2 ml of DPPH solution was mixed with 1 ml of different concentrations (20–100 μ g ml⁻¹ in DMSO) of the compounds. The reaction mixture was mixed and incubated at room temperature in the dark for 30 min. The absorbance was recorded spectrophotometrically at 517 nm. A mixture of 1 ml distilled water and 2 ml DPPH solution was used as the control, and ascorbic acid as the standard. The experiment was performed in triplicate and the results were presented as means \pm standard deviations (mean \pm SD). The percent radical scavenging was calculated by the following equation

DPPH scavenging effect
$$
\% = [(A_c - A_t)/A_c] \times 100
$$

where A_c is the absorbance of the control and A_t is the absorbance of the test compound. IC_{50} values were calculated from the plot of scavenging activity against the concentrations of the samples.

2.6 | Anti-inflammatory screening

The *in vivo* anti-inflammatory activity of the test compounds was performed by the carrageenan‐induced rat The *in vivo* anti-inflammatory activity of the test com-
pounds was performed by the carrageenan-induced rat
paw edema method.^[31] Male Sprague–Dawley rats Fire *in vivo* anti-infiammatory activity of the test compounds was performed by the carrageenan-induced rat paw edema method.^[31] Male Sprague–Dawley rats weighing 160–220 g were divided into groups of six each. The experimental protocol was approved by Institutional weighing 160–220 g were divided into groups of six each.
The experimental protocol was approved by Institutional
Animal Ethics Committee (IAEC/2008/01–11/HSK), and the same procedure was carried at H. S. K. College of Pharmacy, Bagalkot. The hind paw edema was induced Animar Ethes Committee (IAEC/2008/01–11/HSK), and
the same procedure was carried at H. S. K. College of
Pharmacy, Bagalkot. The hind paw edema was induced
in each rat by the sub-plantar injection of 1% λ -carrageenan (0.2 ml in 0.9% NaCl) 1 h after the administration geenan (0.2 fm in 0.9% NaCl) I in after the administration
of the test compounds (5 and 10 mg kg⁻¹ of body weight)
and standard drug (Diclofenac, 10 mg kg⁻¹ of body
weight) orally. The control groups received 0.5% Naand standard drug (Diclofenac, 10 mg kg^{-1} of body in distilled water. The volume of the paw was measured by means of a digital plethysmometer (UGO Basile 7140) at 0.5, 1, 3 and 5 h after injection of the inflammatory stimulus. The percent edema inhibition was calculated by the following equation

%edema inhibition = $[(V_c - V_t)/V_c] \times 100$

where V_c is the edema volume of rat of control group, at time t, and V_t is the edema volume of rat of test compound, at time t.

3 | RESULTS AND DISCUSSION

All the synthesized compounds are air stable and soluble in chloroform, DCM, DMF and DMSO. Analytical data of all the compounds are in good agreement with their proposed molecular formulae. Molecular structures of comm chronorm, DCM, DMF and DMSO. Analytical data of
all the compounds are in good agreement with their pro-
posed molecular formulae. Molecular structures of com-
plexes were finally corroborated by single crystal X-ray diffraction studies. Analytical and spectral parameters are presented in Section 2.

3.1 | Spectral characterization

The numbering scheme of LH_3 is given in Scheme 1. The IR spectrum of the LH_3 (Figure S1) displayed characteristic absorption bands at 3444, 3326, 3232, 1680, 1660 and 16 Humbering scheme of **LH₃** is given in scheme 1. The

1R spectrum of the **LH**₃ (Figure S1) displayed characteris-

tic absorption bands at 3444, 3326, 3232, 1680, 1660 and

1607 cm⁻¹ due to ν (O-H), ν (N1-H), ν (N $(C7 = 01)$, ν $(C14 = 02)$ and ν $(C=N)$, respectively. The band due to $\nu(C7 = 01)$ has remained almost unchanged 1607 cm⁻¹ due to ν (O-H), ν (N1-H), ν (N2-H), ν
(C7 = O1), ν (C14 = O2) and ν (C=N), respectively. The
band due to ν (C7 = O1) has remained almost unchanged
in the spectra of all the complexes, sugge involvement in coordination. The band attributed to ν $(C=N)$ in LH_3 has shifted towards lower frequency upon complexation, indicating the involvement of azomethine nitrogen in coordination. The absence of bands due to ν (C=N) in **LH₃** has simulted towards lower hequency upon
complexation, indicating the involvement of azomethine
nitrogen in coordination. The absence of bands due to ν
(N2-H) and ν (C14 = O2) and the appearance of (N2-H) and ν (C14 = O2) and the appearance of two new bands in the spectra of complexes at about 1611– (N2-H) and ν (C14 = O2) and the appearance of two
new bands in the spectra of complexes at about 1611–
1613 cm⁻¹ and 1318–1324 cm⁻¹ due to the stretching (N2-H) and ν (C14 = O2) and the appearance of two
new bands in the spectra of complexes at about 1611–
1613 cm⁻¹ and 1318–1324 cm⁻¹ due to the stretching
vibrations of the conjugated -C=N-N=C- and enolic The band is in the spectra of complexes at about 1611–1613 cm⁻¹ and 1318–1324 cm⁻¹ due to the stretching vibrations of the conjugated -C=N-N=C- and enolic C14-O2, respectively, indicate the enolization and subsequent coordination of oxygen atom to the central metal ion. In addition, the bands due to ν (C43 = O5) $_{\text{amido}}$ (in the amido form of ligand) and the one due to the new bond in addition, the bands due to ν (C43 = O5)_{amido} (in the amido form of ligand) and the one due to the new bond -C=N-N=C- (in the imidol form of ligand) in complex 4 are appearing at the same frequency (1612 cm^{-1}) ; Figure S2). The IR spectra of isomorphous complexes 5 (Figure S3) and 6 (Figure S4) are nearly identical with minor differences in the lower frequency region. The Figure S2). The IK spectra of isomorphous complexes 5
6 (Figure S3) and 6 (Figure S4) are nearly identical with
minor differences in the lower frequency region. The
band observed in the region of 3420–3560 cm⁻¹ in all complexes was assigned to ν (OH) of coordinated/lattice band observed in the region of 3420–3560 cm⁻¹ in all complexes was assigned to ν (OH) of coordinated/lattice held water molecules. The *tert*-butyl substituent groups in LH_3 and its complexes show their characteristic absorption patterns between 2866 and 2957 cm^{-1} .

The 1 H NMR spectrum of the LH_3 (Figure S5) has shown three singlets at 12.33, 12.16 and 11.13 ppm due to the N2H, N1H and O3H, respectively. But the ${}^{1}H$ NMR spectrum of Ni(II) complex (5) (Figure S6) did not show any signals corresponding to either N2H or O3H, indicating the transformation of the ligand into the imidol form and further deprotonation prior to coordination with the metal ion. In addition, the ligand showed a imalcating the transformation of the figand fino the
imidol form and further deprotonation prior to coordina-
tion with the metal ion. In addition, the ligand showed a
sharp singlet due to an azomethine proton (C15-H) at 8.52 ppm. This signal is being shifted to 8.69 ppm due to the participation of azomethine nitrogen in coordination. The two sharp singlets in $LH_3/5$ at 1.28/0.90 and 1.39/ 6.52 ppm. This signal is being similar to 6.69 ppm due to
the participation of azomethine nitrogen in coordination.
The two sharp singlets in $LH_3/5$ at 1.28/0.90 and 1.39/
1.21 ppm correspond to two sets of magnetically 1.21 ppm correspond to two sets of magnetically non-
equivalent *tert*-butyl groups.^[32] The signals correspond-
ing to the protons of aromatic moieties of LH_3 and 5 were
observed in the range of 7.2–8.25 and 6.73–8. ing to the protons of aromatic moieties of LH_3 and 5 were respectively.

The ¹³C NMR spectrum of LH_3 (Figure S7) has showed signals at 164.32, 164.19 ppm and 155.18 ppm, assigned to carbonyl (C7 and C14) and azomethine (C15) carbons, respectively. The C14 and azomethine (C15) carbons exhibited downfield shifts in the Ni(II) complex (Figure S8). This suggests the involvement of enolic oxygen and azomethine nitrogen in the complexation. The two distinct resonances due to methyl carbons enolic oxygen and azomethine nitrogen in the complexa-
tion. The two distinct resonances due to methyl carbons
 $(-\text{CH}_3)_3$ of two non-equivalent *tert*-butyl groups in LH3/5 occurred at 29.77/29.8 and 31.74/30.79 ppm. The two distinct resonances due to methyl carbons
CH₃)₃) of two non-equivalent *tert*-butyl groups in
 $\frac{1}{3}$ /5 occurred at 29.77/29.8 and 31.74/30.79 ppm.
The ESI-MS of **LH**₃ (Figure S9) shows a molecular ion

The ESI-MS of LH_3 (Figure S9) shows a molecular ion
peak $[LH_3 + H]^+$ at 540. In the positive mode ESI-MS of 4, the base peak observed at 1135 corresponds to mass of $[({\rm Co}({\bf LH})(\mathbf{L}\mathbf{H_2}))+H]^+$. This assignment is in good agree-**4**, the base peak observed at 1135 corresponds to mass of $[(Co(LH)(LH_2)) + H]^+$. This assignment is in good agreement with the ascribed +3 oxidation state for cobalt. ESI-MS analysis of 5 and 6 was done in DMSO with 0.1% NH4OH solution. The aqua ligand present in both the complexes was replaced by $NH₃$. Hence, the peaks at NH_4OH solution. The aqua ligand present in both the complexes was replaced by NH_3 . Hence, the peaks at 613 and 618 in the ESI-MS of 5 and 6 are assigned to $[Ni(\mathbf{L}\mathbf{H})(NH_3) + H]^+$ and $[Cu(\mathbf{L}\mathbf{H})(NH_3) + H]^+$, respectively. In addition, the intense peaks at 674 and 679 in 5 and 6 correspond to the DMSO adducts, i.e. [Ni(LH) $(DMSO) + H$ ⁺ and $[Cu(LH)(DMSO) + H$ ⁺, respec- $(DMSO) + HJ$ and $[Cu(LH)(DMSO) + HJ$, respectively.^[33] The ESI and simulated mass spectra of all the complexes are provided as supplementary information (Figures S10a–S12c). complexes are provided as supplementary information The UV–Vis absorption spectra of an une
plexes are provided as supplementary information
gures S10a–S12c).
The UV–Vis absorption spectra of **LH**₃ and its metal

complexes were recorded in DMF solution and are displayed in Figure S13. LH_3 displays two bands at 266 and 299 nm in the UV region, attributing to $\pi \to \pi^*$ and $n \to \pi^*$ transitions, respectively. After coordination with the metal ions, the n $\rightarrow \pi^*$ transition associated with azomethine chromophore is bathochromically shifted, indicating the involvement of imine nitrogen in coordinathe metal fons, the $n \rightarrow \pi$ transition associated with
azomethine chromophore is bathochromically shifted,
indicating the involvement of imine nitrogen in coordina-
tion.^[34] No d-d transitions could be observed in the of 4 and 5. The intense band that appeared at about marcating the involvement of infine introgen in coordina-
tion.^[34] No d-d transitions could be observed in the case
of **4** and **5**. The intense band that appeared at about
416–442 nm for the complexes has been assigned charge transfer transitions. In the electronic spectrum of 6, the broad band with peak maxima at 666 nm corresponds to the combination of ${}^{2}B_{1g} \rightarrow {}^{2}A_{1g}$ and ${}^{2}B_{1g} \rightarrow {}^{2}E_{g}$ 6, the broad band with peak maxima at 666 nm componds to the combination of ${}^{2}B_{1g} \rightarrow {}^{2}A_{1g}$ and ${}^{2}B_{1g} \rightarrow$ transitions as expected for square-planar geometry.^[35] me broad band with peak maxima at 666 nm corre-
nds to the combination of ${}^{2}B_{1g} \rightarrow {}^{2}A_{1g}$ and ${}^{2}B_{1g} \rightarrow {}^{2}E_{g}$
nsitions as expected for square-planar geometry.^[35]
X-band EPR spectra were recorded in powder

well as in frozen solution of $[Cu(LH)(H₂O)]·H₂O$ (6) in methanol. The EPR signals of polycrystalline samples exhibit isotropic intense broad signals with $g_{iso} = 2.057$ with no hyperfine splitting (Figure S14). From solution EPR measurements, it was possible to resolve the hyperfine pattern (Figure S15) with $g_{\parallel} = 2.245$, $g_{\perp} = 2.034$, g_{av} $= 2.104$, G = 7.206, A \parallel = 154 × 10⁻⁴ cm⁻¹ and A_⊥ = 34 × 10⁻⁴ cm⁻¹. The spectrum shows typical axial behavior with tensor values of $g \parallel > g_{\perp} > g_{e}$ (2.0023), which are consistent with a d_{x-y}^2 ground state.^[36] The axial symmetry parameter, G, quantifies the exchange interaction between copper centers in a polycrystalline compound. It is calculated by using the equation G = g_{\parallel} – 2/g_⊥ – 2. As $G > 4$, it is expected that there is no exchange coupling between two copper centers.[37] The empirical factor $f = g \mid A \mid$ measures the degree of tetrahedral distortion. 6 of 13 WILEY-Organometallic CHIMMALAGI ET AL.

The f value for complex 6 is 146 indicating a small distortion from planarity.[38] According to Kivelson and Neiman,^[39] the ground state molecular orbital coefficient, α^2 , can be taken as a measure of metal-ligand covalency For a value for complex **o** is 140 multeding a small distor-
on from planarity.^[38] According to Kivelson and
eiman,^[39] the ground state molecular orbital coefficient,
, can be taken as a measure of metal–ligand coval and is calculated by the following equation

$$
\alpha^2 = \frac{-A_{\parallel}}{0.036} + \left(g_{\parallel} - 2.0023\right) + \frac{3}{7}(g_{\perp} - 2.0023) + 0.04
$$

The above-calculated value for α^2 (0.72) for 6 is evidence for the partial covalent character of the complex.

3.2 | Thermal analysis

The TG and DT analyses of the crystalline complexes **3.2** | **Thermal analysis**
The TG and DT analyses of the crystalline complexes
were carried out in the temperature range of 25–1000°C under a nitrogen atmosphere. Complex 4 is stable up to 170°C. It undergoes thermal decomposition in three steps were carried out in the temperature range of 25–1000 C
under a nitrogen atmosphere. Complex 4 is stable up to
170°C. It undergoes thermal decomposition in three steps
(Figure S16). The first step occurring at 170–200°C cor sponds to the liberation of hydrogen bonded enclathrated water molecule (found 1.62%, calcd 1.57%). The second (Figure 316). The first step occurring at 170–200 C corresponds to the liberation of hydrogen bonded enclathrated water molecule (found 1.62%, calcd 1.57%). The second step decomposition appears in the range $200-350^{\circ}C$ indicating the decomposition of part of the ligand. The step decomposition appears in the range $200-350^{\circ}\text{C}$, indicating the decomposition of part of the ligand. The last step observed in the range $350-580^{\circ}\text{C}$ corresponds to the decomposition of the remaining part of the ligand. The plateau obtained above 580°C corresponds to the formation of stable cobalt oxide with the residual weight of 7.34%. The TG/DTA curves of 5 (Figure S17) show the first exothermic weight loss (found 2.91%, calcd 2.85%) at 70°C, and is consistent with the removal of hydrogen bonded enclathrated water molecule. In the second stage, coordinated aqua ligand (found 2.93%, calcd 2.85%) is lost for C, and is consistent with the removal of hydrogen
bonded enclathrated water molecule. In the second stage,
coordinated aqua ligand (found 2.93%, calcd 2.85%) is lost
in the range 85–120°C with an exothermic DTA curve a 105°C. The third and fourth step observed in the temperain the range $85-120^{\circ}$ C with an exothermic DTA curve at 105° C. The third and fourth step observed in the temperature range $120-510^{\circ}$ C involve the removal of organic component. The residual weight of 12% above 510°C corresponds to NiO. The TG and DT data of 6 are similar to 5 and are in good agreement with the crystallographic structure.

3.3 [|] Single crystal X‐ray diffraction studies

The crystallographic data of all three complexes are summarized in Table 1. The molecular structures of 4, 5 and 6 along with atom numbering schemes are portrayed in The crystanographic data of an time complexes are sum-
marized in Table 1. The molecular structures of 4, 5 and 6
along with atom numbering schemes are portrayed in
Figures 1–3, respectively. Table 2 lists selected bond lengths and bond angles. Relevant hydrogen bond interactions are compiled in Table S1.

3.3.1 | Structural descriptions of [Co(LH) (LH_2)] $\cdot H_2O$ (4)

The crystallographic asymmetric unit of complex 4 comprises two independent molecules of [Co(LH) $(LH₂)$] (4A and 4B) and two lattice held water of crystallization. These two molecules (4A and 4B) are Comprises two independent indecutes or $[CO(LH)]$
(LH_2)] (4A and 4B) and two lattice held water of crystallization. These two molecules (4A and 4B) are crystallographically non-equivalent and chemically equivalent. The compound crystallizes in orthorhombic system with $P2_12_12_1$ space group, and the Co(III) center exhibits a distorted octahedral coordination where two inequivalent ONO tridentate ligands [that differ in their protonation state $(LH^{2-}$ and LH_2^-); Scheme 2], coordinate metal ions through phenolic oxygen atoms $[(O3a, O5a)$ in **4A**; $(O3b, O5b)$ in **4B**, imine nitrogens [(N3a, N6a) in $4A$; (N3b, N6b) in $4B$], enolic oxygens [O2a in 4A; O2b in 4B] and carbonyl oxygens [O5a in 4A; O5b in 4B]. In both the molecules, the chloro atom present in the ortho position of the 2,4–dichlorophenyl
present in the ortho position of the 2,4–dichlorophenyl group (in LH_2^-) exhibits twofold rotational disorder (Figure S18). The chlorine atoms Cl3a (in 4A) and Cl3b (in 4B) have major occupancy of 0.879(4) and 0.844(5), respectively. In the coordination sphere, both the ligands $(LH²$ and $LH₂⁻)$ in molecule 4A and molecule 4B are almost perpendicular to each other. Azomethine nitrogens [(N3a and N6a) in 4A; (N3b and N6b) in 4B] of the two ligands reside trans to each other, whereas the other two donor sites [(O2a, O5a) and (O3a, O6a) in 4A; (O2b, O5b) and (O3b, O6b) in 4B] have remained *cis* to each other, i.e. the ligands coordinate to the metal in a meridional fashion. The coexistence of both the tautomeric forms of ligand within a complex is substantiated by the bond distances coordinate to the metal in a meridional fashion. The
coexistence of both the tautomeric forms of ligand
within a complex is substantiated by the bond distances
in the region of five-membered chelate rings (Co1A/ N3A/N2A/C14A/O2A and Co1A/N6A/N5A/C43A/O5A in 4A and Co1B/N3B/N2B/C14B/O2B and Co1B/N6B/ In the region of hve-membered cherate rings (C01A)
N3A/N2A/C14A/O2A and Co1A/N6A/N5A/C43A/O5A
in 4A and Co1B/N3B/N2B/C14B/O2B and Co1B/N6B/
N5B/C43B/O5B in 4B). The C14-O2_{imidol} [1.311(5) Å in $NSA/N2A/C14A/C2A$ and C01A/N6A/N5A/C45A/O5A
in **4A** and C01B/N3B/N2B/C14B/O2B and C01B/N6B/
N5B/C43B/O5B in **4B**). The C14-O2_{imidol} [1.311(5) Å in
4A, 1.312(5) Å in **4B**] and C43-O5_{amido} [1.275(5) Å in $M=4A$ and COLB/NSB/NZB/C14B/OZB and COLB/N6B/
N5B/C43B/O5B in **4B**). The C14-O2_{imidol} [1.311(5) Å in
4A, 1.312(5) Å in **4B**] and C43-O5_{amido} [1.275(5) Å in
4A, 1.264(5) Å in **4B**] differ in their lengths. The N5-C43 [1.333(5) Å in **4A**, 1.332(5) Å in **4B**] is more of σ **4A**, 1.312(5) Å in **4B**] and C43-O5_{amido} [1.275(5) Å in **4A**, 1.264(5) Å in **4B**] differ in their lengths. The N5-C43 [1.333(5) Å in **4A**, 1.332(5) Å in **4B**] is more of σ in character compared with N2-C14 [1.304(6) 1.298 (6) Å in **4B**. The bite angles for the ligands $(LH²$ and $LH₂$ ⁻) lie in the range 82.52–96.38°, indicating a distortion from an ideal octahedral geometry, with the *trans*-donor bond angles in the range 173.71–176.94° $\text{Im} \textbf{4}$ A, 1.332(3) A In $\textbf{4}$ B is more of 0
npared with N2-C14 [1.304(6) Å in $\textbf{4}$ A,
 $\textbf{4}$ B]. The bite angles for the ligands
 $\overline{ }$) lie in the range 82.52–96.38°, indicating a distortion from an ideal octahedral geometry, with (LH and LH₂) he in the range 82.52–96.38°, indicating a distortion from an ideal octahedral geometry, with the *trans*-donor bond angles in the range 173.71–176.94° (in **4A**); 175.33–177.73° (in **4B**) and the *cis*-don the *trans*-donor bond angles in the range $173.71-176.94^{\circ}$
(in **4A**); $175.33-177.73^{\circ}$ (in **4B**) and the *cis*-donor bond
angles in the range $82.52-95.92^{\circ}$ (in **4A**); $83.29-96.38^{\circ}$ (in 4B).

The enclathrated water molecules donate hydrogen bonds to enolic and carbonyl oxygen atoms, and receive hydrogen bonds from the amide nitrogen atom (Figure S19). In addition, the molecular structure is bonds to enonc and carbonyl oxygen atoms, and received
hydrogen bonds from the amide nitrogen atoms
(Figure S19). In addition, the molecular structure
stabilized by various C-H $\cdots \pi$ interactions (Figure S20).

TABLE 1 Crystal data and structure refinement details of the complexes 4, 5 and 6

3.3.2 | Structural descriptions of [Ni(LH) (H_2O)]·H₂O (5) and [Cu(LH)(H₂O)]·H₂O (6)

Complexes 5 and 6 are isomorphous structures where different divalent cations are present $[Ni(II)]$ in 5 and $Cu(II)$ in 6]. Both complexes crystallize in the monoclinic system with $P2₁/c$ space group and each asymmetric unit contains neutral $[M(L)(H_2O)]$ (M = Ni, 5; Cu, 6) along with one lattice held water of crystallization. The Ni(II) or Cu(II) centers display a slightly tetrahedrally distorted square-planar coordination provided by two oxygen (O2, O3) and one nitrogen (N3) atoms of the doubly deprotonated ONO tridentate hydrazone (LH^{2−}) and square-planar coordination provided by two oxygen (O2, O3) and one nitrogen (N3) atoms of the doubly deprotonated *ONO* tridentate hydrazone (LH^{2-}) and one oxygen atom (O4) of aqua ligand. The C14-O2 bond deprotonated *ONO* tridentate hydrazone (LH^{2-}) and
one oxygen atom (O4) of aqua ligand. The C14-O2 bond
length [1.311(17) Å in 5, 1.305 (19) Å in 6] is of singlebond character, and C14‐N2 [1.311(19) Å in 5, 1.317 (2) dond character, and C14‐N2 [1.311(19) Å in 5, 1.317 (2) lone oxygen atom (O4) of aqua figand. The C14-O2 bond
length [1.311(17) Å in 5, 1.305 (19) Å in 6] is of single-
bond character, and C14-N2 [1.311(19) Å in 5, 1.317 (2)
Å in 6] is of double-bond character, this indicates enolization and subsequent coordination of oxygen atom

FIGURE 1 ORTEP projection of 4A (drawn at 50% probability level) with partial atom‐numbering scheme. Disordered atoms and H‐atoms attached to carbon omitted for clarity

FIGURE 2 ORTEP projection of 5 showing 50% probability ellipsoids

Media 3 SKIM projection of 6 showing 50% producinty
ellipsoids
M-O bond lengths in 5 and 6 are in the range of FIGURE 3 ORTEP projection of 6 showing 50% probability ellipsoids

(O2) to the central metal ion via deprotonation. The (O2) to the central metal ion via deprotonation. The
ligand has formed one five- and one six-membered (O2) to the central metal ion via deprotonation. The ligand has formed one five- and one six-membered chelate ring with the metal center. The bite angles O2– ligand has formed one five- and one six-membered
chelate ring with the metal center. The bite angles O2-
M-N3 (84.62° in 5; 82.51° in 6) and N3-M-O3 (96.51° in 5; 95.94° in 6) indicate significant deviation from chelate ring with the metal center. The bite angles O2-M-N3 (84.62° in 5; 82.51° in 6) and N3-M-O3 (96.51° in 5; 95.94° in 6) indicate significant deviation from ideal square-planar geometry in the complexes.^[40] The

 $M-O$ bond lengths in 5 and 6 are in the range of 1.809–1.921 Å and 1.860–1.974 Å, respectively. The M-N bond lengths in 5 and 6 are 1.798(12) Å and 1.885(12) Å, respectively. These bond lengths are close 1.809–1.921 Å and 1.860–1.974 Å, respectively. The M-
N bond lengths in 5 and 6 are 1.798(12) Å and
1.885(12) Å, respectively. These bond lengths are close
to the reported analogous square-planar Ni(II) and Cu(II) complexes.[41,42]

TABLE 2 Selected bond lengths (\hat{A}) and angles $(^{0})$ of the complexes **4, 5** and **6**

(Continues)

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WILEY-Organometallic-

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TABLE 2 (Continued)

The molecular structures of 5 and 6 are stabilized by intra‐molecular hydrogen bond interactions, N1‐ H1A \cdots N2. In both 5 and 6, enclathrated water molecule is involved in three different hydrogen interactions (Figures S21 and S22). O5 of lattice water acts as hydrogen bond donor to the carbonyl oxygen (O1) and deprotonated imido oxygen (O2) of two adjacent molecules, and also as hydrogen bond acceptor from oxygen $(O4)$ of aqua ligand. In the case of 5 and 6, and deprotonated finido oxygen (Oz) of two adjacent
molecules, and also as hydrogen bond acceptor from
oxygen $(O4)$ of aqua ligand. In the case of 5 and 6,
only one of the two available O-H bonds of aqua ligand is utilized in the hydrogen bonding, thereby breaking Etter's first rule of hydrogen bonding.^[43] The structures of 5 and 6 are additionally stabilized by various intermolecular $\pi \cdot \pi$ interactions. The $\pi \cdot \pi$ interactions present in 5 and 6 are depicted in Figures S23 and S24, respectively. No significant C-H \cdots π interactions
S24, respectively. No significant C-H \cdots π interactions are observed.

3.4 | Biological studies

3.4 | Biological studies
The stability of the metal complexes (4–6) in DMF and
DMSO was checked by comparing the UV–Vis spectra of the complexes over a period of 5 h (Figure S25). No significant change was observed, which indicates that the complexes are stable in DMF and DMSO for the first 5 h.

3.4.1 | Antioxidant activity

In vitro antioxidant activity of the newly synthesized hydrazone (LH_3) and its complexes was evaluated using the DPPH free radical scavenging assay. In the present investigation, LH_3 and its metal complexes have shown a mydrazone (LH_3) and its complexes was evaluated using
the DPPH free radical scavenging assay. In the present
investigation, LH_3 and its metal complexes have shown a
dose-dependent response (Figure S26). The DPPH scav enging activity of all compounds is expressed as the IC_{50} values. The IC_{50} values of LH_3 , 4, 5 and 6 complexes are 85.76, 42.36, 78.54 and 57.54 ppm, respectively (Table 3). Comparison of the activity exhibited by LH_3 and its complexes indicates that the metal complexes exhibit better activity than the ligands themselves due to their enhanced lipophilicities. Among the complexes, the Co(III) complex exhibited significant DPPH radical scavenging activity. Compared with ascorbic acid taken as a standard drug, the activity shown by reported compounds is less.

3.4.2 | Anti-inflammatory activity

The *in vivo* anti-inflammatory activity of the metal complexes along with parent ligand was evaluated by the func-The *in vivo* anti-inflammatory activity of the metal complexes along with parent ligand was evaluated by the functional model of carrageenan-induced rat paw edema. The *In vivo* and Innanimatory activity of the metal complexes along with parent ligand was evaluated by the functional model of carrageenan-induced rat paw edema.
There are biphasic effects in carrageenan-induced edema.^[44,45] The early phase is mainly mediated by hista-There are biphasic effects in carrageenan-induced edema.^[44,45] The early phase is mainly mediated by histamine, 5-HT, serotonin and kinins^[46], whereas the latter phase is protracted by the release of prostaglandins and leukotriens derived from arachidonic acid^[7], thus the feet rapidly became swollen, reaching close to the control's level by 3 h. The experimental results (Figure S27; Table 4) exhibit the effective inhibitory activity of the synthesized compounds on both phases of carra-Table 4) exhibit the effective inhibitory activity of the synthesized compounds on both phases of carrageenan-induced paw inflammation. All the compounds geenan-induced paw inflammation. All the compounds have shown suppression of edema in a dose-dependent manner. Compared with LH_3 , its metal complexes have exhibited better activity, and this may be due to chelation, which reduces the polarity of the central metal ion because of partial sharing of its positive charge with the donor atoms of the ligand, thus increasing the lipophilic nature of the metal. $[47,48]$ Compared with diclofenac taken as a standard drug, the activity shown by reported compounds is less. LH_3 is active at 10 mg kg^{-1} dose with a percentage inhibition of 85.41%. Among all complexes, 6 has shown promising activity

TABLE 3 DPPH radical scavenging activity of LH₃ and its metal complexes

Results expressed in mean \pm SEM ($n = 3$). ANOVA followed by Tukey's multiple comparison test.

TABLE 4 Anti-inflammatory activity of LH_3 and its metal complexes

	Paw volume in ml (% of edema inhibition)			
Groups	0.5 _h	1 _h	3 _h	5 ^h
Control	1.097 ± 0.044	1.197 ± 0.014	1.170 ± 0.062	1.343 ± 0.028
Diclofenac (10 mg kg^{-1})		$0.030 \pm 0.027***$ (97.26%) $0.030 \pm 0.027***$ (97.49%) $0.108 \pm 0.040***$ (90.77%)		$0.025 \pm 0.013***$ (98.14%)
LH ₃ (5 mg kg^{-1})		0.383 ± 0.082 *** (65.08%) 0.480 \pm 0.098*** (59.90%) 0.687 \pm 0.138*** (41.28%)		$0.483 \pm 0.080***$ (64.03%)
LH₃ (10 mg kg ⁻¹)		$0.160 \pm 0.086***$ (85.41%) $0.370 \pm 0.109***$ (69.08%) $0.653 \pm 0.098***$ (44.18%)		$0.330 \pm 0.080***$ (75.43%)
4 (5 mg kg ⁻¹)		$0.421 \pm 0.034***$ (62.62%) $0.316 \pm 0.053***$ (73.60%) $0.578 \pm 0.055***$ (60.60%)		$0.324 \pm 0.024***$ (75.87%)
4 (10 mg kg^{-1})		$0.331 \pm 0.074***$ (69.82%) $0.247 \pm 0.032***$ (79.36%) $0.833 \pm 0.045***$ (28.80%)		$0.203 \pm 0.004***$ (84.88%)
5 (5 mg kg ⁻¹)		$0.256 \pm 0.038***$ (76.66%) $0.196 \pm 0.026***$ (83.62%) 0.960 ± 0.048 (17.95%)		$0.243 \pm 0.009***$ (81.90%)
5 (10 mg kg ⁻¹)		0.183 ± 0.042 *** (83.31%) 0.130 ± 0.043 *** (89.14%) 0.790 ± 0.041 ** (32.48%)		$0.217 \pm 0.061***$ (83.84%)
6 (5 mg kg^{-1})		$0.276 \pm 0.055***$ (74.84%) $0.250 \pm 0.032***$ (79.11%) 0.917 ± 0.090 (21.62%)		$0.210 \pm 0.017***$ (84.36%)
6 (10 mg kg^{-1})		$0.103 \pm 0.024***$ (90.61%) $0.230 \pm 0.064***$ (80.78%) 0.887 ± 0.033 (24.19%)		$0.194 \pm 0.044***$ (85.55%)

Results expressed in mean \pm SEM ($n = 6$). ANOVA followed by Dunnett's test.

P < 0.01. *P < 0.001 when compared with control group.

with a percentage inhibition of 90.61% in the early with a percentage inhibition of 90.61% in the early
phase of inflammation (0.5 h). Anti-inflammatory activity of the synthesized metal complexes is greater compared with corresponding metal salts (Table S2).

4 | CONCLUSION

4 | **CONCLUSION**
Mononuclear Co(III), Ni(II) and Cu(II) complexes of 2-**4** CONCLUSION

Mononuclear Co(III), Ni(II) and Cu(II) compl

(2,4-dichlorobenzamido)-N'-(3,5-di-*tert*-butyl-2-

hydroxybenzylidene)benzohydrazide were synthesized Mononuclear Co(III), NI(II) and Cu(II) complexes of 2-
(2,4-dichlorobenzamido)-N'-(3,5-di-tert-butyl-2-
hydroxybenzylidene)benzohydrazide were synthesized
and structurally characterized by X-ray crystallography. The molecular structure of complex 4 divulges that both the tautomeric forms of ligand are associated with the Co (III) ion in a meridonial fashion, and adopts a distorted octahedral geometry. While in isomorphous complexes 5 and 6, ligand coordinated to central metal through imidol tautomeric form, and results into distorted square‐planar geometry. The anti‐inflammatory and antioxidant activities of the three complexes and the ligand were evaluated simultaneously. We found that all the compounds showed considerable activity in a concentration‐dependent manner. Among the compounds tested, metal complexes have shown higher activity than parent ligands perhaps due to their enhanced lipophilicities.

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